New Collection and Order for Chlamydia Trachomatis, Neisseria Gonorrhea & Trichomonas Vaginalis

March 7, 2023

This information applies to: *Island Physicians and Nurse Practitioners*

**Replaces February 12, 2023 Communication

The orders for CT/GC PCR and Trichomonas PCR have been combined. The **new order is called CT/GC/TV PCR.** Along with the new order change there are also new collection kits to be used, the BD Molecular Swab Collection kit and the BD Molecular Urine Transport Kit. New collection kits can be ordered on the Provincial Laboratory Supplies Request Form by writing "Chlamydia swabs" or "Chlamydia urines".

	BD Molecular Swab Collection kit	**BD Molecular Urine Transport Kit**
Sample Type	vaginal, cervix/endocervix, throat, and rectal samples*	urine samples* 1st stream urine
Directions	No change to current practices	samples must be collected into a sterile urine container, then transfer **2mL** of urine immediately into the sample buffer tube provided in the collection kit.
Other notes	Please make sure to break off the swab at the marked line when placing it into the sample container, as the swab stick can pierce the lid, resulting in a leak	Do not send the sterile urine container to avoid any confusion for testing. Any urines submitted in urine containers for CT/GC/TV PCR will be rejected as they will be too old for testing.

**Step by step directions are contained on pages 2-3 of this document.*

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Provincial Laboratory Services
Chlamydia/Gonorrhea/Trichomonas PCR Specimen Collection Guide

URINE SPECIMEN COLLECTION PROCEDURE

NOTE: The patient should only be provided with a sterile, plastic, preservative-free specimen collection cup.

1. The patient should not have urinated for at least 1 hour prior to specimen collection.
2. The patient should collect the first 20–60 mL of voided urine (the first part of the stream - not midstream) into a sterile, preservative-free specimen collection cup and securely replace cap.
3. Label the urine collection cup with patient identification and date/time collected.

Transfer of Urine Specimens to the BD Molecular Urine Sample Buffer Tube

First void urine specimens must be transferred from the collection cup to the BD Molecular Urine Sample Buffer Tube immediately after collection. Wear clean gloves when handling the BD Molecular Urine Transport Kit components and specimens. If gloves come in contact with the specimen, immediately change them to prevent contamination of other specimens.

1. Uncap the BD Molecular Urine Sample Buffer Tube and the urine specimen cup.
2. Immediately after collection, use the graduated transfer pipette to mix the urine specimen gently in the collection cup and transfer approximately 2 mL into the BD Molecular Urine Sample Buffer Tube.

NOTE: Use the graduations on the transfer pipette as a guide. DO NOT overfill or under fill the tube.

3. Use the viewing window on the BD Molecular Urine Sample Buffer Tube Label to ensure urine specimen was added to the tube.
4. Discard the transfer pipette in a biohazard waste container.

NOTE: The transfer pipette is intended for use with a single specimen.

5. Tighten the cap securely on the BD Molecular Urine Sample Buffer Tube.
6. Invert the BD Molecular Urine Sample Buffer Tube 3 to 4 times to ensure that the specimen and reagent are well mixed.
7. Label the BD Molecular Urine Sample Buffer Tube with patient information and date/time collected.

NOTE: Do not obscure the barcodes on the tube. Obscuring the barcode may result in instrument errors and inability to test the sample.

8. Transport to the testing laboratory as soon as possible. Specimens in the BD Molecular Urine Sample Buffer Tube can be stored at 2-30°C.

ENDOCERVICAL SWAB SPECIMEN COLLECTION PROCEDURE

1. Remove the sterile swab from its sheath, taking care not to contaminate the tip or shaft. If the swab tip is touched or if the swab is laid down, discard it and use a new collection kit. Check for presence of the swab tip. If the swab has no tip, discard it and request a new BD Molecular Collection Swab.
2. Holding the swab by the cap, insert it into the cervical canal and rotate for 15–30 seconds.
3. Withdraw the swab carefully, avoiding contact with the vaginal mucosa.
4. The swab must be broken into the BD Molecular Swab Sample Buffer Tube immediately after specimen collection. **Proceed directly to the Transfer of Swab Specimens to the BD Molecular Swab Sample Buffer Tube section**

CLINICIAN VAGINAL SWAB SPECIMEN COLLECTION PROCEDURE

1. Remove the sterile swab from its sheath, taking care not to contaminate the tip or shaft. If the swab tip is touched or if the swab is laid down, discard it and use a new collection kit. Check for presence of the swab tip. If the swab has no tip, discard it and request a new BD Molecular Collection Swab.
2. Holding the swab by the cap, insert it into the vagina no more than 2 inches and rotate for 10–15 seconds.
3. Withdraw the swab carefully, avoiding contact with the skin.
4. The swab must be broken into the BD Molecular Swab Sample Buffer Tube immediately after specimen collection. Proceed directly to the Transfer of Swab Specimens to the BD Molecular Swab Sample Buffer Tube section

PATIENT-COLLECTED VAGINAL SWAB SPECIMEN COLLECTION PROCEDURE

NOTE: Ensure that the patient reads and understands the Patient Collection Instructions before providing them with the BD Molecular Collection Swab. The patient should be provided only with the BD Molecular Collection Swab in the sheath.

Patient Instructions

1. Wash hands with soap and water. Rinse and dry.
2. It is important to maintain a comfortable balance during the collection procedure.
3. Remove the sterile swab from its sheath, taking care not to contaminate the tip or shaft. Do not lay the swab down on any surface. If you touch or drop the swab tip or if the swab is laid down, discard it and request a new BD Molecular Collection Swab. Check for presence of the swab tip. If the swab has no tip, discard it and request a new BD Molecular Collection Swab.
4. Hold the swab by the cap in one hand so that the swab tip is pointing toward you.
5. With your other hand, gently spread the skin outside the vagina. Insert the tip of the swab into the vaginal opening. Point the tip toward your lower back and relax your muscles.
6. Gently slide the swab no more than 2 inches into the vagina. If the swab does not slide easily, gently rotate the swab as you push. If it is still difficult, do not attempt to continue. Make sure the swab touches the walls of the vagina so that moisture is absorbed by the swab.
7. Rotate the swab for 10–15 seconds.
8. Withdraw the swab without touching the skin.
9. Replace the swab in its sheath and cap securely.
10. After collection, wash hands with soap and water, rinse and dry.
11. Return the swab in its sheath to the nurse or clinician as instructed

Transfer of Swab Specimens to The Bd Molecular Swab Sample Buffer Tube

Specimens collected using the BD Molecular Collection Swab must be transferred to the BD Molecular Swab Sample Buffer Tube immediately after collection. Wear clean gloves when handling the BD Molecular Swab Collection Kit components and specimens. If gloves come in contact with the specimen, immediately change them to prevent contamination of other specimens.

1. Unscrew the cap of the BD Molecular Swab Sample Buffer Tube, taking care not to contaminate the contents or the outside of the tube.
2. Immediately after collection, insert the BD Molecular Collection Swab into the tube so that the score mark indicated by the black line is at the lip of the tube. Carefully break the swab shaft at the score mark and allow the swab to drop into the tube. Use caution to avoid splashing of the tube contents.
3. Tighten the cap securely on the BD Molecular Swab Sample Buffer Tube.
4. Label the BD Molecular Swab Sample Buffer Tube with patient information and date/time collected.

NOTE: Do not obscure the barcodes on the tube. Obscuring the barcode may result in instrument errors and inability to test the sample.

5. Transport to the testing laboratory as soon as possible. Specimen in BD Molecular Swab Sample Buffer Tube can be stored at 2-30°C.